



STOOLPREPARATION

SEDIMENTATION - METHODE

Material

Stoolpreparation - Sedimentation

- Underlay
- Gloves
- Beaker glass with NaCl 0.9%
- Wooden spatula
- Urine collection cup, conical, plastic
- Strainer (fitting for the collection cup)
- Gauze
- Stool sample (fresh stool)
- Falcon Tubes (15 ml)
- Fresh stool
- examination tubes
- Pasteur pipette + rubber bulb
- Microscope slides + cover slips

Equipment

- Safety cabinet
- Chemical fume hood
- Centrifuge
- Microscope







Providing materials:

Prepare one conical plastic urine collection cup (with sieve and gauze) for each stool sample, as well as one fresh stool examination tube for each sample. Fill the fresh stool examination tubes with NaCl.



- Thoroughly mix and combine a quantity of fresh stool, with NaCl a fresh stool examination tube.
- Filter the prepared stool mixture through a sieve with double-layered gauze into a conical plastic urine collection cup.
- Completely fill the urine collection cup with NaCl solution.

Afterwards, let it stand



- Pour the supernatant into a liquid waste bottle, slowly and continuously, until approximately 245 ml are poured and only the loose sediment remains. Immediately stop pouring at this point.
- Resuspend the sediment thoroughly.
- Prepare two 15 ml Falcon tubes (1x SAF, 1x Native).
- Fill the SAF tube with resuspended sediment.
- Fill the remaining sediment into the Native tube.

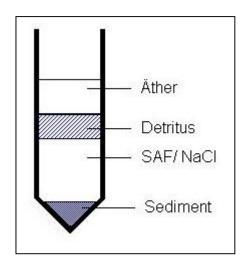


- The sediment from the Native tube is examined "directly."
- Using a glass Pasteur pipette, draw up the sediment.
- Allow the sediment to settle in the pipette for 5 minutes.
- Examine 3 slides with 2 coverslips each, using a 10x objective.
- Fill the second Falcon tube with 10 ml of SAF



- Centrifuge for 1 minute.
- Aspirate the supernatant until reaching the sediment.
- Fill the sediment with NaCl and ether.
- Close the Falcon tube and shake it vigorously.
- Then, centrifuge again.





Aspirate the three upper layers (ether, debris, and NaCl).







- Examine the entire sediment under the microscope using a 10x objective.
- If necessary, examine multiple coverslips.

